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Lot No.: 69V Lot-specific information

Olerup SSP® HLA-A*34

Product number: 101.425-06 – including *Taq* pol.

101.425-06u – without *Taq* pol.

Lot number: 69V

Expiry date: 2016-October-01

Number of tests: 6 Number of wells per test: 10+1

Storage - pre-aliquoted primers: dark at -20°C

PCR Master Mix: -20°C
 Adhesive PCR seals
 Product Insert
 RT

This Product Description is only valid for Lot No. 69V.

Complete product documentation consists of generic Instructions for Use (IFU), lot specific Product Insert, Worksheet and Certificate.

CHANGES COMPARED TO THE PREVIOUS OLERUP SSP® HLA-A*34 Lot (64R)

The HLA-A*34 kit is updated for new alleles to enable separation of:

- Confirmed¹ alleles as listed in the IMGT/HLA database
- Polymorphisms in exons outside of the region encoding the peptide binding domain
- Null and Alternatively expressed alleles

A well containing Negative Control primer pairs has been added.

The format of the Product Insert and Worksheet have been changed.

Three wells have been added to HLA-A*34, wells **9 to 11**.

The HLA-A*34 specificity and interpretation tables have been updated for the HLA-A alleles described since the previous *Olerup* SSP® HLA-A*34 lot was made **(Lot No. 64R)**.

April 2014 Rev. No.: 00

¹As described in section Uniquely Identified Alleles.

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Lot No.: **69V** Lot-specific information

As of lot series V, the Specificity Table is included in the lot-specific Product Insert, and the Interpretation Table is included in the Worksheet.

The primers of the wells detailed below have been exchanged, added or modified compared to the previous lot.

Well	5'-primer	3'-primer	rationale
4	-	Modified	3'-primer modified for increased yield of HLA-specific PCR product.
6	Moved	Moved	Primer pair moved to well 10, for decreased tendency of primer oligomer formation.
9	New	New	New primer pair for the A*34:11 allele.
10	Added	Added	Primer pair added from well 6.
11	-	-	Negative Control.

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Lot No.: 69V Lot-specific information

PRODUCT DESCRIPTION

HLA-A*34 SSP subtyping

CONTENT

The primer set contains 5'- and 3'-primers for identifying the A*34:01 to A*34:11 alleles.

PLATE LAYOUT

Each test consists of 11 PCR reactions in a 16 well cut PCR plate. Wells 12 to 16 are empty.

1	2	3	4	5	6	7	8
9	10	NC	empty	empty	empty	empty	empty

The 16 well cut PCR plate is marked with 'A34' in silver/gray ink.

Well No. 1 is marked with the Lot No. '69V'.

Wells 1 to 10 – HLA-A*34 high resolution primers.

Well 11 - Negative Control (NC).

A faint row of numbers is seen between wells 1 and 2 or wells 7 and 8 of the PCR trays. These stem from the manufacture of the trays, and should be disregarded.

The PCR plates are heat-sealed with a PCR-compatible foil.

Please note: When removing each 16 well PCR plate, make sure that the remaining plates stay sealed. Use a scalpel or a similar instrument to carefully cut the foil between the plates.

INTERPRETATION

Due to the sharing of sequence motifs between HLA-A alleles non-HLA-A*34 alleles will be amplified by primer mixes 1, 2 and 4 to 8.

In addition, a few HLA-B and HLA-C alleles will be amplified by primer mixes 1, 5 and 6.

For further details see Specificity Table.

UNIQUELY IDENTIFIED ALLELES

All the HLA-A*34 alleles, i.e. **A*34:01 to A*34:11 alleles**, recognized by the HLA Nomenclature Committee in January 2014^{1,2} will be amplified by the primers in the HLA-A*34 subtyping kit.

The HLA-A*34 kit enables separation of the confirmed HLA-A*34 alleles as listed in the IMGT/HLA database. An HLA allele is listed as confirmed by IMGT/HLA if it has been sequenced by more than a single laboratory or from multiple sources. Current allele confirmation status for HLA-A*34 alleles is listed below.

The HLA-A*34 kit also enables identification of polymorphisms in exons outside of the region encoding the peptide binding domain and of null and alternatively expressed alleles.



OLERUPSSP®

101.425-06 - including Taq pol., IFU-01 **101.425-06u– without** *Taq* **pol.**, IFU-02

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Lot No.: 69V Lot-specific information

The HLA-A*34 subtyping kit cannot distinguish the silent mutations in the A*34:01:01-34:01:02 or the A*34:02:01-34:02:03 alleles.

¹HLA-A alleles listed on the IMGT/HLA web page 2014-January-17, release 3.15.0,

ALLELE CONFIRMATION STATUS

Allele	Status ¹	Allele	Status ¹
A*34:01:01	Confirmed	A*34:07	Unconfirmed
A*34:01:02	Unconfirmed	A*34:08	Confirmed
A*34:02:01	Confirmed	A*34:09	Unconfirmed
A*34:02:02	Unconfirmed	A*34:10N	Unconfirmed
A*34:02:03	Unconfirmed	A*34:11	Unconfirmed
A*34:03	Confirmed		
A*34:04	Unconfirmed		
A*34:05	Confirmed		
A*34:06	Unconfirmed		

¹Allele status "confirmed" or "unconfirmed" as listed on the IMGT/HLA web page 2014-January-17, release 3.15.0, www.ebi.ac.uk/imgt/hla.

RESOLUTION IN HOMO- AND HETEROZYGOTES

Results file with resolution in HLA-A*34 homo- and heterozygotes is available upon request.

www.ebi.ac.uk/imgt/hla.

Alleles that have been deleted from or renamed in the official WHO HLA Nomenclature up to and including the last IMGT/HLA database release can be retrieved from web page http://hla.alleles.org/alleles/deleted.html.

•LERUPSSP® HLA-A*34

101.425-06 – including *Taq* **pol.**, IFU-01 **101.425-06u– without** *Taq* **pol.**, IFU-02

Visit www.olerup-ssp.com for "Instructions for Use" (IFU)

Lot No.: **69V**

Lot-specific information

SPECIFICITY TABLE

HLA-A*34 SSP subtyping

Specificities and sizes of the PCR products of the 10+1 primer mixes used for HLA-A*34 SSP subtyping

Primer Mix	Size of spec. PCR product ¹	Size of control band ²	Amplified HLA-A*34 alleles ³	Other amplified HLA Class I alleles ⁴
15	100 bp	800 bp	*34:01:01-34:06, 34:08, 34:10N-34:11	*01:13, 01:17, 03:63, 03:88, 11:01:01-11:11, 11:13-11:16, 11:20-11:27, 11:29-11:39, 11:41- 11:52Q, 11:54-11:95, 11:97, 11:99N-11:105, 11:107-11:120, 11:122-11:158, 11:160-11:176, 25:02, 26:13, 26:19, 26:33, 66:01:01-66:01:02, 66:04, 66:06- 66:11, 66:13-66:14, 66:17-66:19, 69:02, C*07:335
2 ⁵	110 bp	1070 bp	*34:01:01-34:01:02, 34:05, 34:11	*26:48, 26:69
3	195 bp	1070 bp	*34:02:01-34:02:03, 34:04, 34:07-34:10N	
4 ⁶	135 bp	800 bp	*34:03, 34:06	*03:01:19, 25:09, 26:14, 26:18, 26:28, 26:73, 31:03-31:04
5	200 bp	800 bp	*34:04	*31:01:07, B*15:82, B*15:260, B*40:186, C*03:186
6	155 bp	1070 bp	*34:05	*02:91, 02:322, 03:94, C*07:81, C*07:243
7	140 bp 215 bp	1070 bp	*34:06 *34:07	*26:18, 31:03-31:04 *11:96
8	200 bp 360 bp	800 bp	*34:08 *34:09	*01:51, 02:55, 03:24, 25:03, 26:20, 32:15, 68:71 *02:135, 02:309, 02:454, 03:01:19, 25:13, 26:30, 26:65, 31:04, 66:02-66:03, 66:12, 66:16
	4751	4070	*0.4.4.4	
9 10	175 bp 185 bp	1070 bp 1070 bp	*34:11 *34:10N	
10 11 ⁷	100 ph	1070 БР	Negative Control	

¹Alleles are assigned by the presence of specific PCR product(s). However, the sizes of the specific PCR products may be helpful in the interpretation of HLA-A*34 SSP typings.

Nonspecific amplifications, i.e. a ladder or a smear of bands, may sometimes be seen. GC-rich primers have a higher tendency of giving rise to nonspecific amplifications than other primers. PCR fragments longer than the control bands may sometimes be observed. Such bands should be disregarded and do not influence the interpretation of the SSP typings.



When the primers in a primer mix can give rise to HLA-specific PCR products of more than one length this is indicated if the size difference is more than 20 base pairs. Size differences of 20 base pairs or less are not given. For high resolution SSP kits, the alleles listed are specified according to amplicon length.

Product Insert Page 6 of 12

101.425-06 – including *Taq* **pol.**, IFU-01 **101.425-06u– without** *Taq* **pol.**, IFU-02

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Lot No.: **69V** Lot-specific information

PCR fragments migrating faster than the control bands, but slower than a 400 bp fragment may be seen in some gel read-outs. Such bands can be disregarded and do not influence the interpretation of the SSP typings.

Some primers may give rise to primer oligomer artifacts. Sometimes this phenomenon is an inherit feature of the primer pair(s) of a primer mix. More often it is due to other factors such as too low amount of DNA in the PCR reactions, taking too long time in setting up the PCR reactions, working at elevated room temperature or using thermal cyclers that are not pre-heated.

²The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 1070 or 800 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the shorter, 800 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

³For several HLA Class I alleles 1st and/or 4th exon(s) and beyond, as well as intron nucleotide sequences, are not available. In these instances it is not known whether some of the primers of the SSP sets are completely matched with the target sequences or not. Assumption is made that unknown sequences in these regions are conserved within allelic groups.

⁴Due to the sharing of sequence motifs between HLA-A alleles non-HLA-A*34 alleles will be amplified by primer mixes 1, 2 and 4 to 8.

In addition, a few HLA-B and HLA-C alleles will be amplified by primer mixes 1, 5 and 6.

⁵HLA-specific PCR products shorter than 125 base pairs have a lower intensity and are less sharp than longer PCR products.

⁶Primer mix 4 may have tendencies of unspecific amplification.

⁷Primer mix 11 contains a negative control, which will amplify more than 95% of HLA amplicons as well as the amplicons generated by control primer pairs. PCR product sizes range from 75 to 200 base pairs. The PCR product generated by the control primer pair is 430 base pairs.

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Lot No.: **69V** Lot-specific information

PRIMER SPECIFICATION

Well No.	1	2	3	4	5	6	7	8	9	10
Length of spec.	100	110	195	135	200	155	140	200	175	185
PCR product							215	360		
Length of int.	800	1070	1070	800	800	1070	1070	800	1070	1070
pos. control ¹										
5'-primer(s) ²	282	270	363	423	78	445	103	102	423	415
	^{5'} -CAg ^{3'}	^{5'} -AAA ^{3'}	^{5'} -ATA ^{3'}	^{5'} -gCT ^{3'}	5' -TCC 3'	5' -TCT 3'	5' -CCT 3'	5' -ACA 3'	^{5'} -gCT ^{3'}	^{5'} -ggT ^{3'}
							423	341		
							^{5'} -gCT ^{3'}	^{5'} -ggC ^{3'}		
3'-primer(s) ³	341	341	517	517	238	559	277	259	559	559
. (,	5' -CgT 3'	^{5'} -CgT ^{3'}	^{5'} -CgT ^{3'}	^{5'} -CgC ^{3'}	5' -CCT 3'	5' -CgT 3'	^{5'} -ggA ^{3'}	^{5'} -gTT ^{3'}	5' -CgC 3'	^{5'} -CgT ^{3'}
							524	418		
							5' -CAC 3'	^{5'} -gTC ^{3'}		
Well No.	1	2	3	4	5	6	7	8	9	10

¹The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 1070 or 800 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the shorter, 800 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

²The nucleotide position matching the specificity-determining 3'-end of the primer is given. Nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

³The nucleotide position matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.



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Lot No.: 69V Lot-specific information

Lot No.: 69V Lot-specific information														
CELL LINE VALIDATION SHEET														
HLA-A*34 SSP subtyping kit ²														
		112/4/	Well											
					1	2	3	4	5	6	7	8	9	10
													Ť	
					201434501	201073902	201073903	201434504	201073905	201434506	201073907	201191008	201434510	201434511
				g	434	073	073	434	073	434	073	191	434	434
				Lot No.:	201	20	201	201	201	201	201	201	201	201
	IHW	/C cell line ¹	A*	A*		-	-	``	``	-				
1	9001		*24:02	, ,	-	-	-	-	-	-	-	-	-	-
2		LK707	*02:01		-	-	-	-	-	-	-	-	-	-
3		E4181324	*01:01		-	-	-	-	-	-	-	-	-	-
4		GU373	*30:01		-	-	-	-	-	-	-	-	-	-
5		KAS011	*01:01		-	-	-	-	-	-	-	-	-	-
6	9353	SM	*02:01	*26:03	-	-	-	-	-	-	-	-	-	-
7	9020		*26:01		-	-	-	-	-	-	-	-	-	-
8	9025		*31:01		-	-	-	-	-	-	-	-	-	-
9	9026		*26:01		-	-	-	-	-	-	-	-	-	-
10	9107	LKT3	*24:02		-	-	-	-	-	-	-	-	-	-
11	9051	PITOUT	*29:02		-	-	-	-	-	-	-	-	-	-
12	9052	DBB	*02:01		-	-	-	-	-	-	-	-	-	-
13	9004	JESTHOM	*02:01		-	-	-	-	-	-	-	-	-	-
14	9071	OLGA	*31:01		-	-	-	-	-	-	-	-	-	-
15	9075	DKB	*24:02		-	-	-	-	-	-	-	-	-	-
16	9037	SWEIG007	*29:02		-	-	-	-	-	-	-	-	-	-
17	9282	CTM3953540	*03:01	*80:01	-	-	-	-	-	-	-	-	-	-
18	9257	32367	*33:03	*74:01	-	-	-	-	-	-	-	-	-	-
19	9038	BM16	*02:01		-	-	-	-	-	-	-	-	-	-
20	9059	SLE005	*02:01		-	-	-	-	-	-	-	-	-	-
21		AMALA	*02:17		-	-	-	-	-	-	-	-	-	-
22		KOSE	*02:01		-	-	-	-	-	-	-	-	-	-
23	9124		*02:01	*34:01	+	+	-	-	-	-	-	-	-	-
24		JBUSH	*32:01		-	-	-	-	-	-	-	-	-	-
25	9049		*33:01		-	-	-	-	-	-	-	-	-	-
26		WT49	*02:05		-	-	-	-	-	-	-	-	-	-
27		CH1007	*24:10	*29:01	-	-	-	-	-	-	-	-	-	-
28		BEL5GB	*02:01	*29:02	-	-	-	-	-	-	-	-	-	-
29	9050		*29:02	+00.0-	-	-	-	-	-	-	-	-	-	-
30	9021		*30:01	*68:02	-	-	-	-	-	-	-	-	-	-
31		DUCAF	*30:02		-	-	-	-	-	-	-	-	-	-
32	9297		*02:01		-	-	-	-	-	-	-	-	-	-
33		MT14B	*31:01		-	-	-	-	-	-	-	-	-	-
34	9104		*31:01		-	-	-	-	-	Ē	-	-	-	-
35		SSTO	*32:01	*11.01	Ι.	-	-	-	-	-	-	-	-	-
36		KT17	*02:06	*11:01	+	-	-	-	-	-	-	-	-	-
37	9065	HHKB	*03:01		-	-	-	÷	÷	-	-	-	-	-
38 39	9099		*02:17 *01:01	*03:01	-	-	-	÷	÷	-	-	-	-	-
40		WHONP199	*02:07	*30:01	Ē	-	H	÷	÷	Ē	H	-	Ē	
41		H0301	*03:01	30.01	Ė	÷	÷	÷	-	÷	÷	÷	Ė	÷
41		TAB089	*02:07		Ē	H	H	÷	÷		H	H	Ē	
42		T7526	*02:07	*02:07		-	H	-	÷	-	-	-		-
44	9076		*66:01	02.07	+	-	H	÷	÷		H		Ē	
44		SHJO	*23:01	*24:02	-	-	÷	÷	÷	-	÷	Ė	<u>-</u>	
46		SCHU	*03:01	24.02	-	-	-	÷	÷	-	-	-	-	
46		TUBO	*02:16	*03:01		-	÷	÷	÷	÷	H	-	E	
48		TER-ND	*02:01	*11:01	-	Ė	Ė	Ė	÷	Ė	Ė	Ė	F	
46	9303	IEK-IND	02.01	11.01	+									

¹The provided cell line HLA specificities are retrieved from the http://www.ihwg.org/hla web site. The specificity of an individual cell line may thus be subject to change.





Product Insert Page 9 of 12

101.425-06 – including *Taq* **pol.**, IFU-01 **101.425-06u– without** *Taq* **pol.**, IFU-02

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Lot No.: **69V** Lot-specific information

²The specificity of each primer solution in the kit has been tested against 48 well characterized cell line DNAs and where applicable, additional cell line DNAs.

No DNAs carrying the alleles to be amplified by primer solutions 4, 5, 7, 9 and 10 were available. The specificities of the primers in primer solutions 4 and 7 were tested by separately adding one additional 5'-primer, respectively one additional 3'-primer. In primer solutions 5 and 9 it were only possible to test the 5'-primers, the 3-primers were not possible to test. In primer solution 10 it was only possible to test the 3'-primers, the 5-primers was not possible to test.

in primer solution 7 one of the 3'-primers was not possible to test. One additional 5'-primer and one additional 3'-primer in primer mix 8 were tested by separately adding one additional 3'-primer or 5'-primer.





Product Insert

Page 10 of 12

101.425-06 – including *Taq* **pol.,** IFU-01 **101.425-06u– without** *Taq* **pol.,** IFU-02

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Lot No.: 69V Lot-specific information





Product Insert

Page 11 of 12

101.425-06 – including *Taq* **pol.,** IFU-01 **101.425-06u– without** *Taq* **pol.,** IFU-02

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Lot No.: 69V Lot-specific information



●LERUPSSP®HLA-A*34 Product Insert Page 12 of 12

101.425-06 – including *Taq* **pol.**, IFU-01 **101.425-06u– without** *Taq* **pol.**, IFU-02

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Lot No.: **69V** Lot-specific information

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